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# Effect of Foliar Application of Gibberellic Acid (GA<sub>3</sub>) and 2,4-Dichlorophenoxyacetic Acid (2,4-D) on Yield and Fruit Quality of Low Seeded Kinnow Mandarin (*Citrus reticulata* Blanco)

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# Abstract

Citrus particularly Kinnow mandarin occupies a prominent position in fruit industry all over the world. Despite a significant increase in Kinnow growing area in Punjab, improvement in fruit quality and yield is still negligible. In order to improve productivity and fruit quality, an experiment was conducted during two successive growing seasons (2012-13 and 2013-14) to investigate the effect of foliar application of the growth regulators (GA<sub>3</sub>, 2,4-D) at pea-sized fruit stage of low seeded Kinnow mandarin plants. The efficacy of GA<sub>3</sub> and 2,4-D were evaluated either in single application or in combinations. The experiment was laid out in Randomized Complete Block Design having eleven treatments and a control, and each treatment was replicated three times. The results revealed that both the growth regulators significantly decreased preharvest fruit drop, leading to improvement in yield and fruit quality in both the seasons. Foliar spray containing 50 mg/L GA<sub>3</sub> + 10 mg/L 2,4-D led to marked increase in fruit weight, diameter, height, volume, shape index, juice percentage, TSS (°Brix), total sugars, and Vitamin C contents compared to control. In addition, significant decrease was observed in pre-mature and mature fruit drop in the treated plants compared to control. These growth regulators in the current concentration can be applied to enhance production and improve citrus fruit quality without compromising on the food safety standards.

#### INTRODUCTION

Citrus is one of the most important fruit crops grown in the tropical and subtropical regions of world. In Pakistan, it is the leading fruit crop with an estimated production of 2.17 million metric tons in a total area of 0.20 million hectares (Ministry of Food and Agric., 2014). Cultural, socio-economic, dietary, nutraceutical and medicinal importance of citrus fruits are well documented (Dugo and Di-Giacomo, 2002; Mabberley, 2004). Various cultivars and varieties are being cultivated in country but Kinnow cultivar is of key importance making Pakistan the largest Kinnow producer in the world. Besides local consumption, Kinnow has high export value due to its eye catching exterior look, excellent internal qualities in terms of taste and flavor, high TSS, juice contents and nutritional value. Pakistan exported 0.393 million metric tons of Kinnow fruits of worth \$ 165.71 million during 2014-15 (Directorate of Agriculture, Punjab, 2015). Currently the share of Pakistan in world citrus fruit exports is about 0.1% which is very low as compared to other exporting countries. In addition, per hectare production of citrus is 9.5 tons which is lower than the potential of 18-20 tons/ha (Ashraf *et al.*, 2015). Among the major causes accounting for low quality and productivity are self-incompatibility, inadequate pollination, nutritional deficiency, water stress, insect-pest and disease infestations and hormonal imbalances (Davies and Albrigo, 1994; Agusti *et al.*, 2006; Alva *et al.*, 2006; Din *et al.*, 2012; Ashraf *et al.*, 2012, 2013).

Citrus fruit plants often produce profuse flowers and consequently a substantial number of flowers and fruits shed off as a way of reducing heavy fruit load (Modise *et al.*, 2009), followed by series of drops during the growth period. There are usually three periods of fruit abscission; the first is the period of fruit set, which usually lasts for a month following full bloom. The second period of intense fruit drop may occur at the onset of hot summer and is referred as 'June drop'. The third period of intense fruit abscission is called as 'preharvest drop' (Racsko *et al.*, 2006). It was reported by Saleem *et al.* (2005) that most of the fruitlets were dropped (80-91%) during the first month after final fruit set. The demand of Kinnow mandarin in local and international markets is increasing day by day. To fulfill this demand, good quality fruit production as well as high production per unit area is required. Improved fruit yield and quality could be obtained by reducing heavy fruit drop (Penter and Stassen, 1999).

Plant growth regulators (PGRs) as foliar applications are the most powerful tools used for manipulating tree growth, flowering, yield and fruit quality traits (Ashraf et al., 2013). In addition, by hastening or delaying fruit maturation, the growers can utilize peak demands, avoid unfavorable environmental conditions and extend the market period (Hegazi, 1980). The application of plant growth regulators can re-enforce hormone balance in the peel, reducing or retarding this precocious fall and the losses at harvest (Almeida et al., 2004). The use of 2,4-dichlorophenoxyacetic acid (2,4-D) and Gibberellic acid (GA<sub>3</sub>) has become a wide spread practice in the citrus producing countries of the world to improve fruit quality and control fruit drop at various stages of fruit growth and development. Exogenous applications of the growth regulators have been tested on different citrus species alone or in combinations (Nawaz et al., 2008; Saleem et al., 2008) either at full bloom or at preharvest stage. The plant growth regulator 2,4-D is playing a vital role in checking pre-harvest fruit drop and ultimately increasing yield without adversely affecting the fruit quality.  $GA_3$  is used widely in various horticultural crops to increase fruit height, diameter, weight and ultimately the yield in mango (Shinde et al., 2008) to stimulate fruit set in peach (Stutte and Gage, 1990) and Clementine mandarin (Talon et al., 1992), and to control cracking of pomegranate fruit (Sepahi, 1986). Moreover,  $GA_3$  has increased the yield of fruit in Balady mandarin (El-Sese, 2005) and soluble solids as well as fruit weight in sweet cherry (Basak et al., 1998). The world health organization (WHO) does not regard 2,4-D and its salts and esters as either genotoxic/carcinogenic and established an acceptable daily intake (ADI) for 2,4-D of 0-0.01 mg/Kg/day (USDA, 2006) which is far less from the concentration applied when it is sprayed as synthetic auxin on fruit trees. The aim of this experiment was to identify foliar sprays of GA<sub>3</sub> and 2,4-D at fruit pea sized stage to increase yield by reducing fruit drop and to improve fruit quality in Low seeded Kinnow under Faisalabad conditions.

### MATERIALS AND METHODS

The present investigation was carried out during two fruiting seasons (2012-13 and 2013-14) on six years old low seeded Kinnow mandarin (Citrus reticulata Blanco) plants grafted on rough lemon (Citrus jambheri Lush) rootstock at citrus orchard, NIAB, Faisalabad, Pakistan. The plants were planted at 21 feet distance under square system, and had similar agro-climatic conditions. Only healthy plants, with uniformity in size, were studied. Before start of the experiment, soil analysis was carried out for confirmation of soil fertility status of the experimental site. The soil was sandy loam, alkaline in reaction (pH 8.1-8.7) with moderate range of available nitrogen and phosphorus. All the treatments were applied in low seeded Kinnow orchard under standard production practices. Aqueous solution of all the treatments was sprayed with a compressed air hand sprayer on whole plants to run off at fruit pea sized stage (5 mm in diameter). The control plants were sprayed with distilled water. The experiment was laid out in randomized complete block design (RCBD) with three replicates for each treatment. The foliar treatments employed were  $T_0$  – Control (sprayed with distilled water only), T<sub>1</sub> - 10 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>2</sub> - 20 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>3</sub> - 30 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>4</sub> - 40 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>5</sub> - 10 mg L<sup>-1</sup> 2,4-D, T<sub>6</sub> - 20 mg L<sup>-1</sup> 2,4-D, T<sub>7</sub> - 30 mg L<sup>-1</sup> 2,4-D, T<sub>8</sub> - 40 mg L<sup>-1</sup> 2,4-D,  $T_9 - 10 \text{ mg L}^{-1} \text{ GA}_3 + 10 \text{ mg L}^{-1} 2,4-D$ ,  $T_{10} - 20 \text{ mg L}^{-1} \text{ GA}_3 + 10 \text{ mg L}^{-1} 2,4-D$ ,  $T_{11} - 30 \text{ mg L}^{-1}$ GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup>2,4-D

# **Data Collection and Evaluations**

At commercial harvest (January), twelve fruits were sampled from all sectors of each replicate to evaluate the treatment effect on fruit quality and yield parameters viz., fruit weight (g), fruit diameter (mm), fruit length (mm), fruit volume (ml), fruit shape index, juice percentage, pH, acidity, TA%, TSS (°Brix), TSS/TA, total sugars, vitamin C contents, number of fruits per plant, fruit weight/plant (Kg), June drop and preharvest fruit drop. Physical and biochemical analysis of fruit were done according to Kassem et al. (2011) at the Citrus Laboratory, NIAB, Faisalabad. Fruit weight was recorded by weighing immediately after harvest. Fruit diameter (mm) and length (mm) were measured with the help of digital caliper. The juice was weighed and expressed as a percentage of the total fruit weight. The TSS (°Brix) of the juice was determined by using hand refractometer (ATAGO, USA Inc). Titratable acidity (TA) expressed as citric acid equivalent was determined by titrating 10 ml of the extracted juice against 0.1 N Sodium hydroxide using phenolphthalein as an indicator. The TSS/TA ratio was determined by dividing TSS with TA value. The vitamin C contents (mg/100 ml juice) were determined by titration with 2, 6 dichlorophenol-indophenol dyes. Total sugars (%) were determined according to Maqbool and Malik (2008). The total number of fruit per plant and weight of fruits/plant (Kg) were determined by harvesting fruits at mid-January in the two experimental seasons.

### **Statistical Analysis**

The current experiments were set up in Randomized Complete Block Design and the data were analyzed using STATISTICA (Version 5.5; Stats Soft). Analysis of variance (ANOVA) was used to test the statistical significance, and the significance of differences among means was carried out using Duncan Multiple Range (DMR) (Duncan, 1955) test at  $P \le 0.05$ .

# **RESULTS AND DISCUSSION**

### Effect on Fruit Weight of Low Seeded Kinnow Mandarin

Table 1 shows data regarding effect of 2,4-D and GA<sub>3</sub> either singly or in combination on fruit weight of low seeded Kinnow. Statistically significant effect of the growth regulators was observed on fruit weight of treated plants compared to untreated control in the current study. We noted that plants treated with 30 mg  $L^{-1}GA_3 + 10$  mg  $L^{-1}$ 2,4-D had the highest fruit weight (174.18 g and 183.33 g) compared to control in both the seasons. In the first experimental season (2012-13), maximum fruit weight (170.15 g) was obtained in 40 mg L<sup>-1</sup> GA<sub>3</sub>, treated plants and in the second year (2013-14), maximum fruit weight (182 g and 177.67 g) was recorded in plants treated with 10 mg  $L^{-1}$  GA<sub>3</sub> + 10 mg L-1 2,4-D and 20 mg L-1 2,4-D, respectively. The current results indicate that an increase in the concentration of GA3 increased the weight of retained fruits. Our results support the findings of Gosh et al. (2015) who reported that the fruit weight was significantly improved due to application of plant growth regulators. Similar enhancement in fruit weight obtained in the present study by the different sprayed substances specially fruit weight and size, and peel weight is also reported in previous experiments (Alberigo, 2002; El-Otmani et al., 2002; Harty et al., 2004). The current results were also in agreement with those of Saraswathi et al. (2003) who observed that 2,4-D and GA<sub>3</sub> and their combinations had beneficial effects on increasing fruit weight. Similarly, the findings of Daulta and Beniwal (1983) are in harmony with the current results who reported that foliar application of growth regulators significantly increased the fruit weight compared to control. Our results are in contrary to the previous report from Brazil which concluded that GA<sub>3</sub> application on 'Monti Parnaso'navel orange trees did not affect the fruit weight (Schafer et al., 2000).

### Effect on Number of Fruits/Plant and Weight of Fruits/Plant (kg)

Foliar applications of 2,4-D and GA<sub>3</sub> on low seeded Kinnow plants had significant effects on number of fruits and weight of fruits per plant compared to control (Table 1). It was noted that the number of fruits and weight of fruits per plant were higher in the second experimental year comparatively to first year results. In 2012-13, the highest number of fruits per plant (514.67) and weight of fruits per plant (89.72 Kg) were obtained from plants treated with 30 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, while in 2013-14 the highest number of fruits per plant (552) and weight of fruits per plant (93.76 Kg) were obtained from plants treated with 20 mg L<sup>-1</sup> GA<sub>3</sub>. The results were found to be in agreement with those of Thomas and Lovatt (2004) and Davies and Zalman (2006) who reported that preharvest application of growth regulators significantly increased number of fruits per plant and ultimately yield. The increase in yield by GA<sub>3</sub> treatments might be due to fact that it stimulates cell division and cell enlargement which reflects on increasing fruit weight and consequently fruit yield (Moore, 1979). Gibberellic acid creates sink strength in the fruit cells, thus attracts water and nutrients, hence increasing fruit weight.

#### Effect on The Physical Characteristics of Low Seeded Kinnow Fruits

It is well established that in citrus with excessive increase in fruit size, the quality is impaired, while on the other side small sized fruits are of low quality. In this study, low seeded Kinnow plants treated with 2,4-D and GA<sub>3</sub> had superior physical fruit characteristics viz. fruit diameter, height, volume, and shape index compared to control (Table 2). Maximum fruit diameter (72.30, 77.84 mm), height (69.96, 61.14 mm), volume (197.35, 202 ml), and shape index (1.01, 0.84) were obtained in treatment of 30 mg L<sup>-1</sup>GA<sub>3</sub>

+ 10 mg L<sup>-1</sup> 2,4-D in both the seasons, respectively. In 2012-13, the highest fruit diameter (72.51 mm) and fruit volume (195.73 ml) were obtained from plants treated with 40 mg L<sup>-1</sup> GA<sub>3</sub> and in 2013-14, the highest fruit diameter (74.43 mm), volume (201 ml) and shape index (0.82) were recorded in 20 mg L<sup>-1</sup> 2,4-D. The use of PGRs in improving citrus fruit quality had been well documented in previous reports (Fidelibus *et al.*, 2002; Saleem *et al.*, 2008, Kassem *et al.*, 2011, Asharf *et al.*, 2012, 2013). GA<sub>3</sub> is well known for its capacity to increase source activity and redistribute carbohydrate, resulting in increased sink strength of developing fruit, either through increased cell division or enhanced cell size (Iqbal *et al.*, 2011). Application of GA<sub>3</sub> and 2,4-D promotes fruit diameter and height as these growth regulators increase plasticity of the cell wall followed by the hydrolysis of starch into sugars. Thus, reduces the cell water potential, resulting in the entry of water into the cell and causing elongation (Tuan *et al.*, 2013). Therefore, spraying GA<sub>3</sub> and 2,4-D greatly accelerating fruit growth rate (height and diameter) in comparison with untreated control.

### Effect on Chemical Characteristics of Low Seeded Kinnow Fruits

The growth regulators (2,4-D and GA<sub>3</sub>) treatments at pea sized fruit stage had an increasing trend towards juice percentage, juice pH, juice acidity, TA%, TSS (°Brix), and total sugars (Table 3, 4). As the concentrations of these growth regulators increased, all these characteristics increased accordingly. Compared to control samples, the highest juice percentage (54.98, 57.3) and pH (3.88, 3.54) were recorded in fruits treated with 30 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D and 40 mg L<sup>-1</sup> 2,4-D, respectively. The increase in juice volume by 2,4-D might be explained by their influence in enhancing cell expansion which increases vesicle capacity for juice accumulation (Agusti et al., 2002). In control fruit samples, juice acidity (7.33 and 8.33%) and TA% (0.92 and 1.06%) were recorded the highest compared to all other treatments. The data on total soluble solids (TSS, °Brix), TSS/TA, vitamin C (mg/100 ml) and total sugars (%) as well as for control samples are presented in Table (4). Statistical analysis indicated that the results are highly significant for all of the above traits in both the seasons. In comparison to control, maximum TSS (11.77, 10.73°Brix), total sugars (10.08, 8.37%) and vitamin C contents (49.2, 44.2 mg/100 ml) were recorded in 30 mg  $L^{-1}$  GA<sub>3</sub> + 10 mg  $L^{-1}$  2,4-D in both the seasons. However, it was observed that these values were higher in the 1<sup>st</sup> experimental year (2012-13) compared to second year (2013-14). The TSS/TA ratio was marginally different among the treatments as well as in the experimental years i.e. it was the highest (15.82) in 40 mg L<sup>-1</sup>GA<sub>3</sub> treated fruits (2012-13) followed by 2013-14 with 13.03. From the current results, we can infer that both 2,4-D and  $GA_3$  had beneficial effect on chemical attributes studied. In case of juice percentage our results were found to be in harmony with Atawia and El-Desouky (1997) and Matthew et al. (2002) who reported that growth regulators sprays at flowering and preharvest stage significantly improved the juice percentage in various citrus species. Results regarding TSS percentage were found to be in pertinent with the report of Huang and Huang (2005) who investigated that application of growth regulators like Auxins and Gibberellins can significantly increase the total soluble solids (TSS) of in citrus fruit juices. The results related to acidity percentage and Vitamin C contents were found to be in close agreement with those of El-Otmani et al. (2004), and Xiao *et al.* (2005). They reported that 2,4-D, GA<sub>3</sub> and NAA significantly reduced acidity percentage, whereas increased the Vitamin C contents of the citrus fruits. The results regarding sugar contents clearly supports the findings of Ingle et al. (2001) and Wang et al. (2004) who reported that 2,4-D and GA<sub>3</sub> and some other growth regulators increased

the total sugar contents in various mandarin and sweet orange cultivars. Increase in TSS might be due to conversion of carbohydrate into simple sugars (Rub *et al.*, 2010) and TSS:TA might be due to increase in TSS and decrease in TA (%) in comparison to control. **Effect on Fruit Drop of Low Seeded Kinnow Plants** 

Foliar application of 2,4-D and GA<sub>3</sub> are regarded as one of the most effective growth regulators in preventing fruit drop in citrus (Michael et al., 1999; Iqbal et al., 2011). The perusal of the Table 5 shows that foliar application of  $GA_{3}$ , 2,4-D and their combinations at different concentrations significantly increased the yield by reducing June and preharvest fruit drop in both the seasons. Preharvest fruit drop is of commercial importance to farmers, as this drop occurs just before harvesting when fruit is physiologically mature. In the current study, the highest June fruit drops (61.01 and 68.58%) as well as preharvest fruit drops (73.26 and 77.67%) were recorded in control plants compared to growth regulator treated Low seeded Kinnow plants. In 2012-13, the lowest June drop and preharvest fruit drops (25.38 and 59.98%, respectively) were obtained in 30 mg  $L^{-1}GA_3 + 10$  mg  $L^{-1}2$ ,4-D treated plants. The current results are in close connection with the findings of Almedia et al. (2004) and Davies and Zalman (2006) who reported that 2,4-D, and GA<sub>3</sub> had significantly reduced fruit drop both in June and at preharvest stage in various citrus species. The results of the present study proved that foliar application of plant growth regulators and their combinations was effective in reducing fruit drop, which ultimately resulted into increase in Low seeded Kinnow fruit yield. The current results also confirm the findings of previous reports which concluded that foliar-applied plant growth regulators were effective in enhancing citrus yield (ElSaida, 2001; Saleem et al., 2005; Omaima and Metwally, 2007; Ashraf et al., 2012, 2013). It seems that GA<sub>3</sub> intensifies an organ ability to function as a nutrient sink and also increases the biosynthesis of IAA in plant tissues which delays the formation of the separation layer therefore enhancing fruit retention (Kassem et al., 2011).

# CONCLUSION

Foliar application of plant growth regulators (GA<sub>3</sub> and 2,4-D) has been a quite common practice in some parts of the world. Improvements in fruit quality and yield and reduction in June and preharvest fruit drop are mostly achieved through exogenous application of plant growth regulators. From current results, we can infer that exogenous application of GA<sub>3</sub> and 2,4-D at fruit pea sized stage markedly increased fruit growth, improved fruit quality and yield by reducing preharvest fruit drop in Low seeded Kinnow plants. Positive results can be achieved, by using the right time of spray and required quantity of both GA<sub>3</sub> and 2,4-D.

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Treat.	Fruit we	Fruit weight (g)		Number of fruits/plant		Fruit weight/plant	
						Kg)	
	2012-13	2013-14	2012-13	2013-14	2012-13	2013-14	
T <sub>0</sub>	132.01h	141.67e	300.33f	364.33e	39.65 g	51.62i	
<b>T</b> <sub>1</sub>	159.06dc	161.67cd	321.33f	493.33c	51.07f	79.78fg	
<b>T</b> <sub>2</sub>	148.84efg	154.67d	467bc	552a	89.72a	93.76a	
<b>T</b> <sub>3</sub>	160.89bcd	168bc	510.33ab	480.67c	82.10b	80.77efg	
$T_4$	170.15ab	145e	441cd	497.67c	75.11bcd	91.18ab	
T <sub>5</sub>	158.84cd	160cd	417de	491.33c	66.28e	78.63 g	
T <sub>6</sub>	157.08cde	177.67ab	459.33cd	527.67b	72.08cde	85.33cde	
<b>T</b> <sub>7</sub>	140.62 gh	170bc	398e	453d	55.88f	76.99 g	
<b>T</b> <sub>8</sub>	164.24bc	168.33bc	483.67abc	515.67b	79.54bc	86.77bcd	
<b>T</b> 9	151.29def	182a	482abc	481.33c	72.91cde	87.62bc	
T <sub>10</sub>	142.12fg	164cd	480.67abc	518b	68.26de	84.9cdef	
T <sub>11</sub>	174.18a	183.33a	514.67a	495c	69.5de	71.78h	
T <sub>12</sub>	166.36abc	166.67c	452.33cd	491c	75.21bcd	81.84defg	

**Table 1:** Effect of growth regulators, on fruit weight, number of fruits/plant and fruit weight/plant of low seeded Kinnow mandarin.

Means in each column followed by the same letters are not significantly different at  $P \le 0.05$  according to Duncan's multiple range test. Treatments: T<sub>1</sub> – 10 mg L <sup>-1</sup> GA<sub>3</sub>, T<sub>2</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>3</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>4</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>5</sub> – 10 mg L<sup>-1</sup> 2,4-D, T<sub>6</sub> – 20 mg L<sup>-1</sup> 2,4-D, T<sub>7</sub> – 30 mg L<sup>-1</sup> 2,4-D, T<sub>8</sub> – 40 mg L<sup>-1</sup> 2,4-D, T<sub>9</sub> – 10 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>10</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>11</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D.

**Table 2:** Effect of growth regulators on physical characteristics of low seeded Kinnow mandarin.

Treat.	Fruit diameter		Fruit	height	Fruit volume (ml)		Fruit shape	
	(mn	n)	(n	<b>ւ</b> m)			index	
	2012-13	2013-	2012-	2013-14	2012-13	2013-14	2012-13	2013-
		14	13					14
T <sub>0</sub>	67.92abcd	67.07d	59.66ab	53.83f	140.89e	152.67e	0.88bcde	0.8abc
<b>T</b> <sub>1</sub>	71.75a	73.48b	59.98ab	57.34de	181.05c	196abc	0.84de	0.78bc
T <sub>2</sub>	69.24abc	72.36b	54.81b	56.74e	157.94d	178.33d	0.79e	0.78bc
T <sub>3</sub>	72.25a	74.38b	69.35a	58.49cde	183.18bc	199.33ab	0.96abc	0.78bc
T <sub>4</sub>	72.51a	69.25cd	67.94a	56.77e	195.73ab	155e	0.93abcd	0.76c
T <sub>5</sub>	70.40ab	73.68b	69.35a	58.4cde	140.48c	187.67bcd	0.91abcde	0.79bc
T <sub>6</sub>	67.44abcd	74.43b	61.43ab	62.41a	176.38c	201a	0.91abcd	0.82ab
T <sub>7</sub>	64.44bcd	72.96b	54.71b	59.88bc	148.39de	192abc	0.85cde	0.82ab
T <sub>8</sub>	68.7abc	73.07b	64.88ab	60.13bc	184.69abc	192.67abc	0.94abcd	0.82ab
T <sub>9</sub>	64.17cd	74.25b	63.33ab	59.51bcd	171.08c	198.33abc	0.99ab	0.82ab
T <sub>10</sub>	62.5d	71.58bc	63.36ab	58.55cde	153.62de	185cd	0.96abc	0.82ab
T <sub>11</sub>	72.30a	77.84b	69.96a	61.14ab	197.35a	202a	1.01a	0.84a
T <sub>12</sub>	67.15abcd	72.6b	64.75ab	59.39bcd	183.54abc	190abcd	0.96abc	0.81ab

Means in each column followed by the same letters are not significantly different at  $P \le 0.05$  according to Duncan's multiple range test. Treatments: T<sub>1</sub> – 10 mg L <sup>-1</sup> GA<sub>3</sub>, T<sub>2</sub> –

 $20 mg L^{-1} GA_3, T_3 - 30 mg L^{-1} GA_3, T_4 - 40 mg L^{-1} GA_3, T_5 - 10 mg L^{-1} 2,4-D, T_6 - 20 mg L^{-1} 2,4-D, T_7 - 30 mg L^{-1} 2,4-D, T_8 - 40 mg L^{-1} 2,4-D, T_9 - 10 mg L^{-1} GA_3 + 10 mg L^{-1} 2,4-D, T_{10} - 20 mg L^{-1} GA_3 + 10 mg L^{-1} 2,4-D, T_{11} - 30 mg L^{-1} GA_3 + 10 mg L^{-1} 2,4-D, T_{12} - 40 mg L^{-1} GA_3 + 10 mg L^{-1} 2,4-D, T_{12} - 40 mg L^{-1} GA_3 + 10 mg L^{-1} 2,4-D, T_{10} - 20 mg L^{-1} 2,4-D, T_{10} - 20 mg L^{-1} 2,4-D, T_{10} - 20 mg L^{-1} GA_3 + 10 mg L^{-1} 2,4-D, T_{10} - 20 m$ 

Treat.	Juice percentage		Juice	рH	Juice Acidity		Titratable acidity	
					(%)		(TA) (%)	
	2012-13	2013-	2012-	2013-	2012-	2013-	2012-13	2013-14
		14	13	14	13	14		
T <sub>0</sub>	50.23d1	52.93ab	3.58c	3.4a	7.33a	8.33b	0.92a	1.06ab
$T_1$	50.76cd	55.37ab	3.68bc	3.49a	5.77b	6.53c	0.74b	0.84cd
$T_2$	54.73a	54.3ab	3.73abc	3.73a	5.3b	7.17bc	0.68b	0.92bcd
T <sub>3</sub>	52.26abcd	48.87b	3.63c	3.41a	5.77b	7.7bc	0.74b	0.98bc
$T_4$	50.9cd	53.77ab	3.72abc	3.5a	5.4b	9.87a	0.69b	1.23a
<b>T</b> <sub>5</sub>	52.66abcd	55.43ab	3.71abc	3.44a	5.47b	6.5c	0.70b	0.83cd
<b>T</b> <sub>6</sub>	51.24bcd	55.37ab	3.6c	3.48a	5.73b	6.07c	0.73b	0.78d
<b>T</b> <sub>7</sub>	54.14abc	57.27a	3.72abc	3.47a	5.9b	6.3c	0.76b	0.81cd
T <sub>8</sub>	54.15abc	54.8ab	3.88a	3.54a	5.10b	6.73c	0.65b	0.86cd
<b>T</b> 9	50.60d	53.57ab	3.71abc	3.54a	6.23ab	6.2c	0.80ab	0.79cd
T <sub>10</sub>	46.61e	56.93ab	3.87a	3.44a	5.83b	6.93bc	0.75b	0.89bcd
T <sub>11</sub>	54.98a	57.3a	3.77abc	3.42a	6.0ab	7.5bc	0.78ab	0.96bcd
T <sub>12</sub>	54.59ab	55ab	3.86ab	3.42a	6.03ab	7.33bc	0.72b	0.94bcd

**Table 3:** Effect of different growth regulators on juice quality characteristics of low seeded Kinnow mandarin.

Means in each column followed by the same letters are not significantly different at  $P \le 0.05$  according to Duncan's multiple range test. Treatments: T<sub>1</sub> – 10 mg L <sup>-1</sup> GA<sub>3</sub>, T<sub>2</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>3</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>4</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>5</sub> – 10 mg L<sup>-1</sup> 2,4-D, T<sub>6</sub> – 20 mg L<sup>-1</sup> 2,4-D, T<sub>7</sub> – 30 mg L<sup>-1</sup> 2,4-D, T<sub>8</sub> – 40 mg L<sup>-1</sup> 2,4-D, T<sub>9</sub> – 10 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>10</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>11</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>14</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>15</sub> – 40 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>5</sub> + 10 mg L<sup>-1</sup> 2

**Table 4:** Effect of growth regulators on chemical characteristics of low seeded Kinnow mandarin.

Treat.	Total soluble solids (TSS) (°Brix)		TSS	5/TA	Vitamin C (mg/100 ml)		Total sugars (%)	
	2012-	2013-	2012-	2013-	2012-	2013-	2012-13	2013-
	13	14	13	14	13	14		14
T <sub>0</sub>	11.0b <sup>1</sup>	10.03 b	13.52b	9.73bc	39.2cd	36.8ab	9.04abc	6.82b
T <sub>1</sub>	11.33ab	10.07b	15.24ab	12.1ab	45.6ab	37.4ab	8.17bcd	8.57a
T <sub>2</sub>	11.37ab	10.27ab	17.16a	11.23ab	41.8bc	37.7ab	9.15abc	7.29ab
T <sub>3</sub>	11.23ab	10.47ab	14.50ab	11.07ab	34.67d	36.73ab	8.93abc	7.39ab
$T_4$	11.13ab	10.33ab	15.82ab	8.23c	41.6bc	34.67b	6.83d	8.24ab
T <sub>5</sub>	10.90b	10.3ab	15.68ab	12.73a	35.8d	41.47ab	7.62cd	7.9ab
T <sub>6</sub>	11.0b	10.03b	15.04ab	12.57a	39.23cd	37.47ab	9.47ab	8.27ab
T <sub>7</sub>	11.30ab	10.5ab	15.10ab	13.03a	41.77bc	41.07ab	9.17abc	8ab

T <sub>8</sub>	11.03b	10.43ab	16.97a	12.14ab	39.6cd	36.43ab	8.46abc	8.25ab
T <sub>9</sub>	11.07ab	10.17ab	13.93b	12.81a	40.93bc	41.37ab	9.0abc	8.32ab
T <sub>10</sub>	11.27ab	10.47ab	15.12ab	11.94ab	48.1a	37ab	8.76abc	7.64ab
T <sub>11</sub>	11.77a	10.73a	14.29ab	11.11ab	49.2a	44.2a	10.08a	8.37ab
T <sub>12</sub>	11.03b	10.27ab	15.37ab	10.95ab	47.43a	38.3ab	8.39abcd	7.77ab

Means in each column followed by the same letters are not significantly different at  $P \le 0.05$  according to Duncan's multiple range test. Treatments: T<sub>1</sub> – 10 mg L <sup>-1</sup> GA<sub>3</sub>, T<sub>2</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>3</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>4</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>5</sub> – 10 mg L<sup>-1</sup> 2,4-D, T<sub>6</sub> – 20 mg L<sup>-1</sup> 2,4-D, T<sub>7</sub> – 30 mg L<sup>-1</sup> 2,4-D, T<sub>8</sub> – 40 mg L<sup>-1</sup> 2,4-D, T<sub>9</sub> – 10 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>10</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>11</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>14</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>15</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>16</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2

**Table 5**: Effect of growth regulators on June and pre-harvest fruit drop of Low seeded Kinnow mandarin.

Treatments	June fru	it drop (%)	Pre-harvest f	ruit drop (%)
	2012-13	2013-14	2012-13	2013-14
T <sub>0</sub>	61.01a <sup>1</sup>	68.58a	73.26ab	77.67a
T <sub>1</sub>	55.24a	59.33bcd	73.22ab	62.33de
T <sub>2</sub>	53.79ab	65.67ab	72.27ab	72.33ab
T <sub>3</sub>	37.01cd	53.83def	66.72bc	58.08ef
$T_4$	51.57ab	63.17abc	74.18ab	66.58bcd
T <sub>5</sub>	42.10bc	59.08bcd	69.42abc	65.42cd
T <sub>6</sub>	53.22ab	65.42ab	73.32ab	70.08bc
T <sub>7</sub>	52.60ab	58.33bcde	74.12ab	66bcd
T <sub>8</sub>	56.79a	48.42f	77.95a	52.75fg
T9	29.11de	57.17bcde	61.52cd	63.17de
T <sub>10</sub>	53.76ab	47.83f	71.23ab	48.67 g
T <sub>11</sub>	25.38e	55.5cdef	59.98d	68.5bcd
T <sub>12</sub>	54.76a	50.42ef	75.78ab	50.42 g

Means in each column followed by the same letters are not significantly different at  $P \le 0.05$  according to Duncan's multiple range test. Treatments: T<sub>1</sub> – 10 mg L <sup>-1</sup> GA<sub>3</sub>, T<sub>2</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>3</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>4</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub>, T<sub>5</sub> – 10 mg L<sup>-1</sup> 2,4-D, T<sub>6</sub> – 20 mg L<sup>-1</sup> 2,4-D, T<sub>7</sub> – 30 mg L<sup>-1</sup> 2,4-D, T<sub>8</sub> – 40 mg L<sup>-1</sup> 2,4-D, T<sub>9</sub> – 10 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>10</sub> – 20 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>11</sub> – 30 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D, T<sub>12</sub> – 40 mg L<sup>-1</sup> GA<sub>3</sub> + 10 mg L<sup>-1</sup> 2,4-D.